
***EFFECT OF SOME ESSENTIAL OILS
ON SOME PATHOGENIC HUMAN'S MOUTH BACTERIA***

By

Ashraf R. El-Zainy
Home Economics Dept.,
Faculty of Specific Education,
Mansoura University, Egypt

Mona Y. Mostafa
Home Economics Dept.,
Faculty of Specific Education,
Mansoura University, Egypt

Research Journal Specific Education

Faculty of Specific Education

Mansoura University

ISSUE NO. 73 JANUARY , 2023

**EFFECT OF SOME ESSENTIAL OILS
ON SOME PATHOGENIC HUMAN'S MOUTH BACTERIA**

*Ashraf R. El-Zainy**

*Mona Y. Mostafa***

Abstract:

Herbal remedies are now associated with safety, in contrast to synthetics, which are seen to be harmful to persons and the environment. This investigation aimed to test in vivo the effectiveness of herbs essential oils on oral bacteria. The antibacterial activity of myrrh (*Commiphora molmol*), thyme (*Thymus vulgaris* L.), sage (*Salvia officinalis* L.), chamomile (*Matricaria recutita* L.) and green tea (*Camellia sinensis*) essential oils (5 mg oil/100 ml water) dilution against total bacterial counts in saliva was studied. The counts of *Streptococcus mutans* and *Lactobacillus rhamnosus* were investigated. A standard susceptibility agar dilution method was used to assess the minimal inhibitory zones (MIZ) of myrrh, thyme, sage, chamomile, and green tea essential oils on *Streptococcus mutans* and *Lactobacillus rhamnosus* in order to look into their potential as antibacterial agents. Data were statistically processed. The experimental group showed a statistically significant reduction in colony counts of *Streptococcus mutans* and *Lactobacillus rhamnosus* relative to the control group. These findings showed the efficacy of the tested herbs oils against halitosis and oral flora, achieving a balance between the harmful and the harmless oral bacteria and also providing a promising practical application for the development of customised, all-natural antibacterial remedies.

Keywords: *Streptococcus mutans* - *Lactobacillus rhamnosus* – Halitosis- Myrrh – Thyme – Sage - Chamomile - Green tea - Essential oil.

Introduction

Herbal mouthwashes are in increasing demand since they target oral infections and provide immediate pain relief while having fewer adverse

* Home Economics Dept., Faculty of Specific Education, Mansoura University, Egypt

** Home Economics Dept., Faculty of Specific Education, Mansoura University, Egypt

effects. Hydrogen peroxide and chlorhexidine, which are found in chemical mouthwash, instantly whiten, sterilise, and reduce tooth discomfort. However, they can discolour teeth and have adverse effects, but they are inexpensive (**Banu and Gayathri, 2016**).

It is generally known that oral conditions can affect a person's quality of life and can cause systemic and antecedent conditions. Many oral problems are strongly influenced by microorganisms. There is a constant demand for alternative therapies that are inexpensive, efficient, and non-toxic, such as plant and herbal medicines, due to the rise in bacterial resistance to antibacterial agents, the toxic side effects of some common antibiotics, and the cost of therapy (**Palombo, 2011**).

Halitosis is defined as foul breath caused by a range of factors, including but not limited to periodontal disease, bacterial coating of the tongue, systemic illnesses, and various foods (**American Academy of Periodontology, 2001**). The exhaled air contains odoriferous chemicals that are linked to the aetiology of halitosis, particularly volatile sulphur compounds (VSC) produced by bacteria (**Rösing and Loesche, 2011**).

Calculus, plaque, and infrequent dental visits were all linked to severe halitosis in a research conducted in Sweden by **Söder *et al.*, (2002)**. Periodontitis and tongue coating was linked to VSC scores in a Japanese study by **Miyazaki *et al.*, (1995)**. In addition, patients with severe periodontitis had greater halitosis scores than non-periodontal patients. Tongue coating was regarded as an influencing factor for halitosis in two Swiss investigations by **Bornstein *et al.*, (2009a)** and **Bornstein *et al.*, (2009b)**. Higher halitosis scores were linked to smoking and periodontal disease (**Bornstein *et al.*, 2009a**). While **Yokoyama *et al.* (2010)** reported that halitosis has been linked to plaque buildup and tongue covering.

Gingivitis/periodontitis can produce odoriferous substances due to the presence of bacteria and inflammatory products. The presence of gingivitis or periodontitis was linked to halitosis in cross-sectional studies (**Quirynen *et al.*, 2009**). The capacity of probable periodontal pathogens and inflammatory products to create volatile odoriferous chemicals was

proven in vitro and in vivo studies (**Yoneda et al., 2010; Salako and Philip, 2011**). More over 60% of the 2000 patients at the breath clinic who had tongue coating, whether it was present alone or in conjunction with periodontal inflammation, had halitosis, according to **Quiryneen et al. (2009)**. The coating on the back of the tongue has been implicated in most investigations, It is in line with the fact that there are billions of bacteria there, including anaerobes, which can make odoriferous substances (**Kazor et al., 2003**). In vitro, bacteria from saliva, plaque removed from gingivitis/periodontitis and the tongue (**Kazor et al., 2003**) create odoriferous compounds, according to **Takeshita et al., (2010)**. These bacteria are reduced in intervention studies that provide a clinically relevant benefit in decreasing halitosis (**Shinada et al., 2010 ; Fedorowicz et al., 2008**). **Silveira et al. (2011)** demonstrated that in people with periodontitis, robust supragingival plaque management can lower VSC and organoleptic ratings. Several studies have shown that halitosis can be reduced by eliminating germs on the tongue's dorsum. One of the most essential approaches for halitosis, according to a study, is tongue cleansing (**Faveri et al., 2006**). Because halitosis is caused by the presence of microorganisms from oral biofilms, every therapeutic method that affects the oral microbiota has the potential to reduce halitosis. Mouthwashes, particularly chlorhexidine and cetilpyridinium chloride, have been shown to reduce halitosis (**Shinada et al., 2010**).

The α -haemolytic streptococcal species *Streptococcus mutans* and *Lactobacillus rhamnosus* are the primary bacterial aetiological agents in caries. (**Taylor et al., 2005**). *Streptococcus mutans* (SM) is the most common bacteria in dental plaque and is the cause of caries (**Matalon et al., 2009**)

Many efforts have been made to exploit medicinal plants by increasing the number of strains resistant to antibacterial chemicals. Herbs have been used in traditional medicine all throughout the world since prehistoric times (**Hose, 2002**).

The stem of *Commiphora molmol* (Family: Burseraceae), a tree that thrives in the Arabian Peninsula and northeast Africa, yields myrrh, an oleo gum resin (Massoud *et al.*, 2001). In folk medicine, it is used for a range of diseases and is referred to as "a pharmacy unto itself." Myrrh has therapeutic properties against a variety of ailments, including antibacterial and anti-inflammatory properties (Kumari *et al.*, 2011 and Cheng *et al.*, 2011). As an antibacterial and anti-inflammatory, myrrh is used to treat tonsillitis, throat infections, and mouth ulcers. It's also utilised at funerals and as a perfume scent (Van Wyk and Wink, 2004). *Staphylococcus aureus* and other gram-positive and gram-negative bacterial species displayed effective antibacterial action when myrrh was utilized. Additionally an astringent, myrrh calms inflamed mouth and throat tissues. Research on the anticancer and analgesic effects of myrrh resin is ongoing (Al-Harbi *et al.*, 1994 and Dolara *et al.*, 1996). Topical myrrh is used to treat mild oral and pharyngeal mucosal inflammations (Sheir *et al.*, 2001).

Thyme extract mouthwash was successful as an antimicrobial agent. When compared to a strong antiseptic like chlorhexidine, it greatly decreased the total number of bacteria in children's saliva (Abdel Hameed *et al.*, 2020). A gargle of sage tea is widely prescribed to cure a sore throat, mouth inflammations, and gingivitis in modern European herbal medicine. Sage is used to treat mouth, throat, and tonsil inflammation (ESCOP, 1996). To treat oral herpes, apply a salve comprised of thyme, myrrh, and goldenseal. Thyme can also be used to treat halitosis and persistent candidiasis (Weiss, 1988).

German Chamomile (GC) is an anti-inflammatory, antibacterial, bacteriostatic, wound-healing promoter, and deodorant that has been used as mouthwash or dentifrice with other herbal ingredients to reduce plaque formation and improve gingival health (Gultz *et al.*, 1998). *Matricaria Chamomilla* is a well-known component in alternative medicine. Dermatological, gastrointestinal, neurological, and mental conditions have all been treated with it (Blumenthal *et al.*, 2003). Oral hygiene has been demonstrated to improve by using Chamomile as a toothpaste or an oral rinse (Pistorius *et al.*, 2003). A specific ingredient in the mouthrinse may be

responsible for the anti-inflammatory properties of chamomile extract, which have been the subject of numerous research. In addition to being used in some toothpastes to treat oral irritations and minor infections, chamomile is also used as a mouthwash to treat gingivitis and periodontal disease (Nissen *et al.*, 1998)

Green tea stops bacteria from multiplying and causing tooth decay. Chamomile oil has antibacterial, anti-fungal, and anti-inflammatory effects. Chamomile oil also aids in the formation of granulation and epithelization. Thyme oil is used as an antibacterial and antiseptic agent against a variety of pathogens. Thyme oil can also help to reduce inflammation (D'Amelio and Mirhom, 2010). Anecdotal observations in the Japanese literature, such as "those who drink a big amount of green tea on a regular basis have less tooth decay" (Kubo *et al.*, 1992) and "drinking green tea keeps the mouth clean" (Sakanaka *et al.*, 1989). Kaneko *et al.* (1993) discovered that a four-week mouth washing routine with a weak catechin solution reduced periodontal disease-related halitosis; it was later shown that tea catechins deodorised methyl mercaptan, the principal source of halitosis (Yasuda and Arakawa, 1995).

Aim of work: The present investigation was carried out to study the antibacterial activity of myrrh, thyme, sage, chamomile and green tea essential oils (5mg/100 ml water) dilution against oral bacterial counts, *Streptococcus mutans* and *Lactobacillus rhamnosus*.

Materials and Methods

Materials

Myrrh (*Commiphora molmol*), thyme (*Thymus vulgaris* L.), sage (*Salvia officinalis* L.), chamomile (*Matricaria recutita* L.), green tea (*Camellia Sinensis*) were purchased from local market and their essential oils were obtained from the National Research Center, Giza, Egypt.

Media: Mitis Salivarius Bacitracin Agar (MSBA), Mann Rogosa Sharpe agar (MRSA) and Nutrient Agar (NA) were obtained from Beta Company for chemicals in Mansoura city, Egypt.

Strains of oral bacteria: The bacterial strains used in this study were *S. mutans* ATCC 25175 and *Lactobacillus. rhamnosus* ATCC 7469 were obtained from Cairo Microbiological Resources Center (MIRCENT), Faculty of Agriculture, Ain Shams University, Egypt and checked for purity in laboratory.

Volunteers: Sixty healthy volunteers ranging in age from 16 to 20 years from Mansoura University students. Ethical standards were upheld, and authorization was requested from the relevant department, Faculty of Specific Education, Mansoura University.

Methods:

Total phenolic compounds were assessed using the method of **Waskmundzka *et al.* (2007)** published in Food Technology Research Institute Agric. Rec. Cent., Egypt.

Identification of herbs essential oils by GC: The volatile oils were separated using a Ds chrom 6200 Gas Chromatograph equipped with a flame ionisation detector, as described by **Singh *et al.*, (2007)**.

Experimental design of Saliva:

Sixty healthy volunteers were recruited and randomly divided into six groups ($n=10$):

* **Group (A)** participants were asked to rinse their mouths with (40 mL) of a placebo mouthwash (control).

* **Group (B)** participants were asked to rinse their mouths with (40 mL) of diluted myrrh essential oil (5 mg/100 ml water), for 1 minute, three times a day for a week.

* **Group (C)** participants were asked to rinse their mouths with (40 mL) of diluted thyme essential oil (5 mg/100 ml water), for 1 minute, three times a day for a week.

* **Group (D)** participants were asked to rinse their mouths with (40 mL) of diluted sage essential oil (5 mg/100 ml water), for 1 minute, three times a day for a week.

* Group (E) participants were asked to rinse their mouths with (40 mL) of diluted chamomile essential oil (5 mg/100 ml water), for 1 minute, three times a day for a week.

* Group (F) participants were asked to rinse their mouths with (40 mL) of diluted green tea essential oil (5 mg/100 ml water), for 1 minute, three times a day for a week.

Saliva samples were collected after 7 days.

Enumeration of total bacterial count (TBC): The total microbiological count in saliva was determined using plate count agar medium (A.P.H.A., 1971). 1.00 ml saliva was put to agar medium-coated Petri-plates, which were then incubated at 37°C for 48 hours. The total bacterial count was calculated using a logarithmic transformation of the average CFU/ml.

Enumeration of *Streptococcus mutans* and *Lactobacillus rhamnosus*:

They were determined using *Lactobacillus rhamnosus* selective MRS agar (Himedia) for *Lactobacillus rhamnosus* screening and *Mitis salivarius* bacitracin agar (MSB) for *Streptococcus mutans* under aerobic incubation at 37°C for 3 days, according to the manufacturer's instructions (Badet and Quero., 2011). The dilution of Herbs essential oils was 5 mg/100ml water. The results are presented as a logarithmic transformation of the average CFU/ml.

Determination of minimum inhibition zones (MIZ)

According to Sagdic, *et al.* (2002), the paper disc technique was used to test the sensitivity of oral bacteria to the selected herbal essential oils. Sterile paper discs (What man No 1, 6.0 mm in diameter) were soaked in essential oils and serially diluted. (5ml oil/100ml water) was the dilution utilised. The discs were then placed on the surface of the inoculated plate count agar medium with *Streptococcus mutans* and *Lactobacillus rhamnosus*.

Statistical analysis:

According to **McCormick and Salcedo (2017)**, all tests were done using the computer application of statistical analysis programme (SPSS, version 24).

Results and discussion

Table 1 shows the total phenols and flavonoid contents (*mg/100g*) of the tested herbs. Total phenols were recorded (114.8, 941.7, 1916.2, 58.9 and 79.4 *mg/100ml*) for myrrh, thyme, sage, chamomile and green tea, respectively. Flavonoid contents recorded (87.6, 154.6, 1360.1, 716.4 and 37.2 *mg/100ml*) for myrrh, thyme, sage, chamomile and green tea, respectively. Phenolic acids and flavonoids have been identified as typical phenolics with an antioxidant action (**Kahkönen et al., 1999**). The total phenolic content of myrrh oil was discovered to be (3.3 percent). Myrrh oil has a total flavonoid concentration of 0.2 per cent (**Mahboubi and Kazempour, 2016**). The resin of the species *C. myrrha* has 44.77 per cent total phenolics (**Danial and Majrashi, 2016**). Total phenolic contents of thyme were 783.81 mg/l, while total phenolic contents of sage were 122.98 mg/l, according to **Viuda-Martos et al. (2010)**. Sage has a total phenolic content of 27.94 mg GAE/100g DW and a total flavonoid content of 27.54 mg/CE/100 mg DW (**Atanassova et al., 2011**). Green tea extract with approximately 50% polyphenols, chlorophyll KK, oregano oil, peppermint oil, clove bud oil, lavender oil, thyme oil, cinnamon bark oil, eucalyptus oil, and mixtures there of **D'Amelio and Mirhom (2010)** found that each of these ingredients can be combined in various proportions to get the necessary antibacterial and anti-inflammatory characteristics in the final mixture. Green tea's chemical makeup is a muddled mess. Polyphenols, specifically flavonoids like catechins, catechin gallates, and proanthocyanidins, are the compounds in green tea that are most predominant (**Graham, 1992**).

In Table 2, the GC results of myrrh (*Commiphora molmol*) essential oil show that curzerene is the major constituent (40.0%) followed by Furanoeudesma-1,3-diene, β -elemene then 2-O-acetylen-1,8,12-epoxy-

germacra-1(10),4,7,11-tetraene, isomer with percentage of 4.5, 8.0 and 6.0%, respectively. When evaluated separately, a number of these chemicals showed strong antibacterial activity, according to several studies (**Ben Arfa et al., 2006; Bassolé et al., 2010 and Bajpai et al., 2012**). **Chen et al. (2013)** found that the highest percentage component in myrrh essential oil was (12.01%) 2-cyclohexen-1-one, 4-ethynyl-4-hydroxy-3,5,5-trimethyl, followed by -elemene, copaene, and aromadendrene (6.18, 5.50 and 4.62 % , respectively). In contrast, new components such as Coaene and -amorphenone were found in myrrh oil (5.50 and 1.96 % , respectively). Other components of Ethiopian species include furanodiene (19.7%), furanoeudesma-1,3-diene (34.0%), and lindestrene (12.0%) (**De Rapper et al., 2012**)

The essential oils of the thyme (*Thymus vulgaris L.*) were examined by GC and represented in Table (3). Thymol is the major constituent (46.3%) followed by p-Cymene, γ -Terpinene then Carvacrol (23.2, 18.3 and 1.5%, respectively). These results agreed with those of **Asllani and Toska (2003)** as they reported that the primary components found were thymol (21.38–60.15 %), p-cymene (7.76–43.75 %), -terpinene (4.20–27.62 %), carvacrol (1.15–3.04 %), and -caryophyllene (1.30–3.07 %). The terpene phenols thymol and carvacrol, according to **Stahl-Biskup (1989)**, are the most significant chemicals in the genus, followed by linalool, p-cymene, -terpinene, borneol, terpinen-4-ol, and 1, 8-cineol. Terpinen-4-ol (13.1 %), -terpinene (9.2%), cis-sabinene hydrate (7.6%), linalool (7.1 percent), and p-cymene (5.7 percent) were the primary ingredients in thyme essential oil (**Viuda-Martos et al., 2010**). Thymol, carvacrol, p-cymene, -terpinene, linalool, -myrcene, and terpinen-4-ol are the primary constituents. Some chemicals are glycosides in part (**Committee on Herbal Medicinal Products, 2013**). The phenolic components of *Thymus* and *Origanum* species, such as thymol and carvacrol, have been linked to high antibacterial activity (**Sokovi et al., 2009 and Hazzit et al., 2009**). Essential oils' antibacterial action is dependent on their chemical ingredients, according to **Boruga et al. (2014)**. The content of phenolic chemicals (thymol) and terpene hydrocarbons (γ -terpinene), respectively, appears to be connected to the antibacterial activity of the essential oil studied (**Boruga et al., 2014**).

However, p-cymene has been linked to synergistic effects with thymol and -terpinene, which could be another explanation for the antibacterial activity found. However, multiple studies have found that essential oils have more potent antibacterial properties than their primary constituents or mixtures, indicating the synergistic effects of minor constituents and the significance of each component in connection to EOs biological activity (**Gill et al., 2002; Rota et al., 2008 and Dorman and, Deans, 2000**).

In Table (4), the GC results of sage (*Salvia officinalis* L.) essential oil show that α -Thujone is the major constituent (35.6%) followed by camphor, α -pinene and β - Thujone (23.0, 9.7 and 8.6%, respectively). According to **Craft (2017)**, The sage oils were primarily composed of the monoterpenoids thujone (17–27%), 1,8-cineole (12–27%), and camphor (13–21%), with low amounts of thujone (3.8–6.0%), camphene (3.5–5.3%), and the sesquiterpene humulene (3.1–4.4%). This chemical profile is similar to many sage oil descriptions previously reported by (**Jirovetz et al., 2006; Alizadeh and Shaabani, 2012; Seidler-Ło'zykowska et al., 2015**). Camphor (25.0%), 1,8-cineole (24.7%), and camphene were the predominant components in sage essential oil (7.6 percent). The most abundant compounds in clove essential oil were eugenol (85.5%) and β -caryophyllene (10.5%) (**Viuda-Martos et al., 2010**). The findings of **MITIĆ-ĆULAFIĆ et al. (2005)** show that thujone, among the primary sage monoterpenes, is harmful to all microorganisms examined. The antibacterial activity of EO and fractions, on the other hand, is unrelated to their amount of thujone, cineole, or camphor, implying that their antibacterial impact is likely due to a synergistic interaction of numerous elements. According to **Selim et al. (2022)**, the GC study of sage (*Salvia officinalis*) EO revealed 21 different components, accounting for 89.94% of the total oil component. 1,8-cineole (39.18 %), β -caryophyllene (12.8 %), and α -terpineol were the most common chemicals (10.3 %).

In Table (5), the GC results of chamomile (*Matricaria recutita* L.) essential oil show that α -bisabolol is the major constituent (46.4%) followed by Terpinen-4-ol (22.1%), β -bisabolol (7.3%) then Viridiflorene (6.1%). These findings are consistent with those of **Afify et al. (2012)**, who found

that α -bisabolol oxide A (35.251 %) and trans- α -fararsene are the most abundant essential oils in chamomile (7.758 %). Bisabolol oxides, bisabolone oxide, α -bisabolol, spathulenol, enyne-dicycloethers, and chamazulene were shown to be the most physiologically active chemicals in chamomile oil, according to **Orav et al. (2010)**. Meanwhile, **Raal et al. (2012)** discovered that chlorogenic acids, ferulic acid glycosides, dicaffeoyl quinic acids, and apigenin glycosides were the predominant phenolic components in chamomile infusions.

In Table (6), the GC/MS results of green tea (*Camellia Sinensis*) essential oil show that terpinen-4-ol is the major constituent (30.3%) followed by β -Linalool, γ -terpinene and α -terpinene (13.3, 10.8 and 10.0%, respectively). On the other hand, hexadecanoic acid, heneicosane, transgeraniol, and nerolidyl acetate are the primary components of GT essential oil, according to **Hu et al. (2010)**, accounting for 30.2 %, 7.36 %, 7.02 %, and 4.7 %, respectively. The strongest bactericidal effects were observed in EOs containing terpene alcohols, followed by those of aldehydes or phenols, such as cinnamaldehyde, citral, carvacrol, eugenol, or thymol. Other EOs, including -myrcene, -thujone, or geranyl acetate, have significantly less effects when combined with ketones or esters. Terpene hydrocarbon-containing volatile oils were primarily inactive (**Barros et al., 2009 and Ait-Ouazzou et al., 2011**).

Sensory evaluation of the tested herb's volatile oils on rinsing are recorded in Table (7). Results show that *mouthwashes* with diluted essential oils (5 mg/100 ml water) of myrrh, thyme, sage, chamomile and green tea were chosen to be the best ones according to the sensory evaluation as they recorded the highest scores for taste, odor, texture color and overall acceptability as compared to (10 and 15) mg/100 ml. It could be noticed that these sensory parameters decreased significantly ($p < 0.5$) by increasing the herbs oil concentration in *mouthwash*. Myrrh is utilised in spices, skin ointments, and fragrances because of its essential oil, and its tincture is used to treat oropharyngeal inflammations (**Brieskorn et al., 1982**). Thyme is a single species herbaceous plant that grows in hilly locations and is used as a beverage instead of or in addition to tea. It is also used to some foods to give

them a pleasing flavour. The plant is commonly used in folk medicine, where it is given to heal oral infections (**Mohamed et al., 2013**). Sage has been used in the food industry as a flavouring ingredient (**Gali-Muhtasib et al., 2000**). German chamomile flower heads have been used to make extracts and herbal beverages. German chamomile flower heads and extracts are used in a variety of herbal medicines, herbal teas, cosmetics, food tastes, dyes, and pest repellents (**Singh et al., 2011**). Tea is one of the most popular beverages in many cultures, second only to water (**Pastoriza et al., 2017**). Its peculiar flavour, scent, and health-promoting properties, as well as its socio-cultural associations, are highly loved around the world (**Komes et al., 2010**).

Data represented in Table 8 showed that rinsing with the tested herbs Eos (5% concentration) significantly ($p < 0.5$) controlled the total microbial counts in saliva compared to the control group. Saliva of the myrrh EO mouthwash group recorded the lowest total microbial counts (2.69 ± 0.06 log cfu/ml), followed by the saliva of the thyme EO mouthwash group (2.75 ± 0.27 log cfu/ml), saliva of sage EO mouthwash group (3.11 ± 0.04 log cfu/ml), saliva of chamomile EO mouthwash group (3.30 ± 0.06 log cfu/ml), and saliva of green tea EO mouthwash group (3.36 ± 0.04 log cfu/ml). A significant decrease in saliva total microbial counts was observed in the tested herbs groups comparing with the control group which recorded the highest total microbial count (4.20 ± 0.04 log cfu/ml). Myrrh tincture is listed in the British Herbal Pharmacopoeia (**BHMA 1996**) as a mouthwash for gingivitis and ulcers. Myrrh was approved by the European Commission (**Blumenthal et al. 2000**) for topical treatment of mild oral and pharyngeal mucosa inflammation. Myrrh has recently been discovered to have antibacterial properties against resistant forms of *S. aureus*, *S. enterica*, and *K. pneumoniae* (**Rahman et al., 2008**). The greater antifungal activity of the myrrh extract film corresponds to the ayurvedic practice of using myrrh extract for oral and vaginal hygiene, antiseptic action, and as a natural antibiotic (**Treadway, 1998**). Additionally, *S. officinalis* EO at 5% concentration shown outstanding in vitro inhibitory action toward the biofilm formation of various *S. enterica* isolates, according to **Selim et al.**

(2022). Ferrazzano *et al.* (2011) found that a green tea extract was effective against cariogenic oral bacteria. The antibacterial impact of the herbal mouth rinse was most likely the reason for its success. Antimicrobial effects of goldenseal were found in oral pathogens such as *S. mutans* and *Fusobacterium. nucleatum* (Pourabbas *et al.*, 2005).

Inhibitory effect of tested herbs volatile oils on oral bacteria log cfu/ml) of saliva groups shown in Table 9. Significant ($p < 0.5$) differences were observed between the control and herbs EO (5% concentration) mouthwash groups in *Streptococcus mutans* and *Lactobacillus rhamnosus* counts. Rising with herbs EOs controlled the oral bacteria in saliva groups. The lowest *Streptococcus mutans* count was for saliva of the myrrh EO mouthwash group (2.0 ± 0.05 log cfu/ml), followed by saliva of the sage EO mouthwash group (2.30 ± 0.17 log cfu/ml), saliva of chamomile EO mouthwash group (2.69 ± 0.15 log cfu/ml), and saliva of green tea EO mouthwash group (2.77 ± 0.22 log cfu/ml), then saliva of thyme EO mouthwash group (2.84 ± 0.10 log cfu/ml). Meanwhile, *Lactobacillus rhamnosus* counts were recorded (2.77 ± 0.12 , 2.69 ± 0.09 , 2.95 ± 0.04 , 2.84 ± 0.04 and 2.90 ± 0.05 log cfu/ml) for the same saliva groups, respectively. The control group recorded the highest *Streptococcus mutans* and *Lactobacillus rhamnosus* counts in saliva (3.30 ± 0.11 and 3.11 ± 0.03 log cfu/ml) respectively. In dentistry, herbal extract is utilized as a teeth cleanser, anti-inflammatory, antibacterial, and analgesic. Because of their safety and low cost, herbal remedies are used by 80 percent of the world's population for basic health care, according to the WHO. *Commiphora myrrha* is used in dentistry to treat bad breath and as an anti-inflammatory in periodontitis (Kumar *et al.*, 2013). Antimicrobial activity of thyme essential oil against oral bacteria such as *Staphylococcus aureus*, *Streptococcus mutans*, *Lactobacillus spp.*, *Pseudomonas aeruginosa*, and *Proteus spp.* was studied (Al- Mahdy *et al.*, 2021). On oral bacteria strains, the effects of thymol, a plant-derived antibacterial agent, were investigated. The relationship between thymol leakage producing concentrations and minimal inhibitory and bactericidal concentrations suggests that membrane perforation is a major mechanism of action for this drug (Shapiro and

[Guggenheim, 1995](#)). The antibacterial properties of plant oils were evaluated against oral germs; the most potent essential oils were Australian lea tree oil, peppermint oil, and sage oil, with thymol and eugenol as potent essential oil components ([Shapiro et al., 1994](#)). Catechins are inhibitory for *Streptococcus. mutans* and *Streptococcus. sobrinus*, according to several researchers, with MICs ranging from 50 to 1000 mg/ml, well within the quantities observed in brewed tea ([Sakanaka et al. 1989](#); [Kawamura and Takeo 1989](#); [Rasheed and Haider 1998](#)).

Zones of growth inhibition (*mm*) of tested herb's volatile oil components (5% concentration) on oral bacteria are registered in Table 10. *Streptococcus mutans* zone growth values were recorded (10, 8, 9, 7 and 8 *mm*), while *Lactobacillus rhamnosus* zone growth values recorded (8, 10, 10, 6 and 8 *mm*) for myrrh EO, thyme EO, sage EO, chamomile EO and green tea EO, respectively. It is noticeable that myrrh EO was more effective in controlling *Streptococcus mutans* growth, followed by sage EO, thyme EO, green tea EO and chamomile EO. Meanwhile, thyme and sage essential oils were more effective in *Lactobacillus rhamnosus* growth inhibition, followed by myrrh and green tea EOs then chamomile EO. Oral ulcers, gingivitis, sinusitis, glomerulonephritis, brucellosis, and a range of skin ailments are treated with myrrh, which possesses antibacterial properties ([Shin et al., 2019](#)). An interesting antimicrobial activity of myrrh extract was demonstrated by [Bechecker et al. \(2022\)](#), especially in relation to Gram-negative bacteria. The inhibition zones range from 16 to 30 mm, while the MIC values range from 15.62 to 250 *mg/ml*. The most effective against *Streptococcus mutans* was a 1 percent solution of thyme essential oil in ethanol, which might be deemed practical as a toothpaste ingredient, both in terms of cost and sensory profile. Furthermore, tests of the formulation's features revealed that the product is stable ([Gonçalves et al., 2011](#)). The inhibitory impact of direct exposure was studied using the plate counting approach. Against the bacteria tested, all of the Thyme essential oils tested demonstrated considerable bacteriostatic activity. Against gram-positive bacteria, its action was more evident ([Marino and Bersani, 1999](#)). The number of *Streptococcus mutans* in tooth plaque was successfully reduced

by using Sage mouthwash (**Beheshti-Rouy et al., 2015**). According to **Ahmed et al. (2017)**, green tea and chamomile tea drinking reduces salivary pH and inhibits salivary *Streptococcus mutans* count. In impoverished nations like India, a green tea mouth rinse can be an effective preventive home treatment. In addition, **Neturi et al. (2014)** discovered that rinsing with green tea reduced the number of *S.mutans* in plaque when compared to the gold standard mouthwash as a control.

Table (1): Total phenol and flavonoid mg/100g contents of dry herbs

Herbs essential oils	Total phenol mg/100ml	Flavonoid mg/100ml
Myrrh	114.8	87.6
Thyme	941.7	154.6
Sage	1916.2	1360.1
Chamomile	58.9	716.4
Green tea	79.4	37.2

Table (2): Volatile components of myrrh essential oil.

Compounds	%	Compounds	%
Curzerene	40.0	<i>cis</i> - β -elemenone	0.9
Furanoeudesma-1,3-diene	14.5	γ -cadinene	0.7
β -elemene	8.0	β -caryophellene	0.7
2-O-acetyle1-8,12-epoxy-germacra-1(10),4,7,11-tetraene, isomer	6.0	β -bourbonene	0.6
2-O-methyl-8,12-epoxy-germacra-1(10),4,7,11-tetraene, isomer	4.0	9- <i>epi</i> -caryophellene	0.5
γ -eudesmol	2.5	α -Humulene	0.4
γ -elemene	2.2	γ -Muurolene	0.3
7- <i>epi</i> - α -eudesmol	2.0	Elemol	0.2
Alloaromadendrance	1.9	2-hydroxyfuranodiene	0.2
Furanodiene	1.3	Dehydroaromadendrance	0.1

Table (3): Volatile components of thyme essential oil.

Compounds	%	Compounds	%
Tymol	46.3	Terpineol	0.9
p-Cymene	23.2	α -Pinene	0.9
γ -Terpinene	18.3	Linalool	0.8
Caravcol	1.5	Sabinene	0.8
α -Terpinolene	1.4	Camphene	0.7
β -Myrcene	1.3	1,8-Cineol	0.5
α -Terpinene	1.1	Geranic acid	0.3
α - Terpineol	1.1	Citral	0.2
4-Terpineol	1.0	Anisole	0.2
α -Thujone	1.0	Geraniol	0.1

Table (4): Volatile components of sage essential oil.

Compounds	%	Compounds	%
α -Thujone	35.6	β -Caryophyllene	0.9
Camphor	23.0	β -Pinene	0.8
α -Pinene	9.7	Tricyclene	0.7
β - Thujone	8.6	α -Terpinolene	0.5
Camphene	7.9	γ -Terpinene	0.4
1,8-Cineole	5.9	Terpinene-4-ol	0.4
Limonene	1.7	α - Terpinene	0.2
α -Humulene	1.6	Thymol	0.1
Borneol	1.4	α -Phellandrene	0.1
Myrcene	1.2	Myrtenol	0.1

Table (5): Volatile components of chamomile essential oil.

Compounds	%	Compounds	%
α -bisabolol	46.4	Trans-nerolidol	1.0
Terpinen-4-ol	22.1	Guaiazulene	0.6
β -bisabolol	7.3	Spathulenol	0.5
Trans-trans-farnesol	6.6	α -phellandrene	0.4
Viridiflorene	6.1	Cis- β -farnesene	0.3
β -bisabolene	2.1	Chamazulene	0.3
α -cubebene	1.9	Methyl acetate	0.3
α -pinene	1.4	Sabinene	0.3
α -bisabolol oxide A	1.4	T-terpinene	0.3
Caryophyllene oxide	1.2	β -pinene	0.1

Table (6): Volatile components of green tea essential oil.

Compounds	%	Compounds	%
terpinen-4-ol	30.3	α - pinene	2.0
β -Linalool	13.3	p-Cymene	2.0
γ -terpinene	10.8	Nerol	1.7
α - terpinene	10.0	1,2-Epoxylinool	1.4
trans-Geraniol	6.8	1,8- Cineole	1.2
Heneicosane	6.5	Geranyl acetone	1.0
Nerolidyl acetate	4.9	1 β -Cadin-4-en-10-ol	0.9
α -Terpineol	3.2	n-Docosane	0.8
α -terpinolene	2.3	n-Pentacosane	0.5
6-Pentadecen-1-ol	2.1	Solanone	0.1

Table (7): Sensory evaluation of mouthwashes with herbs essential oils.

Essential oils Samples	Taste (10)	Odor (10)	Texture (10)	Color (10)	Overall acceptability (10)
5% Myrrh	8.4±0.15a	8.7±0.35a	8.5±0.38a	8.9±0.15a	8.6±0.23a
10% Myrrh	8.0±0.60ab	8.4±0.43ab	8.1±0.60ab	8.5±0.29ab	8.2±0.33a
15% Myrrh	7.4±0.31b	8.0±0.41a	7.5±0.41a	7.9±0.69b	7.6±0.26b
5% Thyme	8.2±0.44a	8.0±0.76a	8.5±0.29a	8.8±0.57a	8.3±0.16a
10% Thyme	7.9±0.85a	7.5±0.55ab	8.0±0.39ab	7.7±0.51b	7.7±0.54b
15% Thyme	7.3±0.25a	7.0±0.36b	7.8±0.25b	7.2±0.38b	7.3±0.21b
5% Sage	8.6±0.47a	8.0±0.79a	8.2±0.31a	8.5±0.45a	8.3±0.33a
10% Sage	8.0±0.72ab	7.9±0.72a	7.8±0.60a	8.3±0.25a	8.0±0.36a
15% Sage	7.6±0.38b	7.2±0.58a	7.1±0.29b	8.0±0.79a	7.4±0.15b
5% Chamomile	8.1±0.60a	8.4±0.15a	8.0±0.79a	8.2±0.44a	8.1±0.16a
10% Chamomile	7.5±0.41a	7.4±0.29b	7.6±0.38a	7.2±0.67b	7.4±0.02b
15% Chamomile	6.4±0.29b	6.8±0.60b	6.1±0.69b	6.9±0.34b	6.5±0.09c
5% Green tea	7.4±0.29a	7.8±0.58a	7.5±0.76a	7.5±0.73a	7.5±0.07a
10% Green tea	7.2±0.60a	7.2±0.67ab	7.6±0.93a	7.2±0.75a	7.3±0.55a
15% Green tea	6.5±0.38b	6.6±0.44b	6.3±0.60a	7.1±0.29a	6.6±0.28b

Table (8): Total microbial evaluation (log cfu/ml) of saliva groups after rinsing with herbs EOs (5% concentration).

Groups TBC	Group A (control)	Group B (Myrrh EO)	Group C (Thyme EO)	Group D (Sage EO)	Group E (Chamomile EO)	Group F (Green tea EO)
Saliva log cfu/ml	4.20±0.04 ^a	2.69±0.06 ^d	2.75±0.27 ^d	3.11±0.04 ^c	3.30±0.06 ^b	3.36±0.04 ^b

EO:essential oil TBC: Total bacterial count

Table (9): Inhibitory effect of herbs EOs (5% concentration) on oral bacteria (log cfu/ml) of saliva groups.

Groups Oral bacteria	Control	Myrrh EO	Thyme EO	Sage EO	Chamomile EO	Green tea EO
<i>Streptococcus mutans</i>	3.30±0.11 ^a	2.0±0.05 ^c	2.84±0.10 ^b	2.30±0.17 ^c	2.69±0.15 ^b	2.77±0.22 ^b
<i>lactobacillus rhamnosus</i>	3.11±0.03 ^a	2.77±0.12 ^{bc}	2.90±0.05 ^b	2.69±0.09 ^c	2.95±0.04 ^b	2.84±0.04 ^{bc}

EO:essential oil

Table (10): Zones of growth inhibition (mm) showing antibacterial activity for herbs EOs (5% concentration) on oral bacteria.

Groups Oral bacteria	Control	Myrrh EO	Thyme EO	Sage EO	Chamomile EO	Green tea EO
<i>Streptococcus mutans</i>	0.0	10	8	9	7	8
<i>lactobacillus rhamnosus</i>	0.0	8	10	10	6	8

EO:essential oil

Conclusion

Daily use of an antimicrobial mouth rinse that has been shown to have strong antibacterial action against *Streptococcus mutans* and *Lactobacillus rhamnosus* would be a good complement to mechanical oral hygiene treatments and could be a valuable component of oral hygiene

regimens. In the prevention and treatment of many human infections, phytopharmaceuticals based on an examined essential oil from myrrh, thyme, sage, chamomile, and green tea may be suitable.

References

- **Abdel Hameed RG, Mostafa MH and El-Malt MA (2020):** Evaluation of the Antimicrobial Effect of Thyme Extract on *Streptococcus Mutans*. *Al-Azhar Dental Journal for girls*,7 (2-A):313-318.
- **A.P.H.A (1971):** American Public Health Association. Recommended method for the microbiological examination of foods. Amer. *Public health Association*, Inc., New York.
- **Afify, A; Ali, F S and Turky, A F (2012):** Control of *Tetranychus urticae* Koch by extracts of three essential oils of chamomile, marjoram and Eucalyptus. *Asian Pacific Journal of Tropical Biomedicine*, 24-30.
- **Ahmed M, Pavani B, Tanzila T, Thanga G, Thejaswini B, Viha P, Udha B and Vaishnavi N (2017):** Effects of green tea and chamomile tea on plaque pH, salivary pH, *Streptococcus mutans* count. *Indian Journal of Pharmaceutical and Biological Research*.5(4).
- **Ait-Ouazzou A, Cherrat L, Espina L, Lorán S, Rota C and Pagán R (2011):** The antimicrobial activity of hydrophobic essential oil constituents acting alone or in combined processes of food preservation. *Innov. Food Sci. Emerg.*, 12: 320–329.
- **Al-Harbi MM, Qureshi S, Raza M et al. (1994):** Anticarcinogenic effect of *Commiphora molmol* on solid tumors induced by Ehrlich carcinoma cells in mice. *Chemotherapy*, 40: 337–347.
- **Alizadeh A and Shaabani M (2012):** Essential oil composition, phenolic content, antioxidant and antimicrobial activity in *Salvia officinalis* L. cultivated in Iran. *Adv. Environ. Biol.*, 6: 221–226.
- **AL-Mahdi Z, Ubaid I and Witwit LJ (2021):** Activity of Cloves, Cinnamon and Thyme Essential Oils Against Some Oral Bacteria. DOI:[10.32441/kjps.05.01.p2](https://doi.org/10.32441/kjps.05.01.p2).
- **American Academy of Periodontology (2001):** Glossary of Periodontal Terms. 4th ed. Chicago: American Academy of Periodontology, 56 p.

- [Asllani U](#) and [Toska V](#) (2003): Chemical Composition of Albanian Thyme Oil (*Thymus vulgaris* L.). *Journal of Essential Oil Research* , 15(3).
- [Atanassova M](#), [Georgieva S](#) and [Ivancheva K](#) (2011): TOTAL PHENOLIC AND TOTAL FLAVONOID CONTENTS, ANTIOXIDANT CAPACITY AND BIOLOGICAL CONTAMINANTS IN MEDICINAL HERBS. *Journal of the University of Chemical Technology and Metallurgy*, 46 (1): 81-88.
- [Badet C](#) and [Quero F](#) (2011): The in vitro effect of manuka honeys on growth and adherence of oral bacteria. *Anaerobe*, 17: 19-22.
- [Bajpai, VK](#), [Baek, KH](#) and [Baek SC](#) (2012): Control of Salmonella in foods by using essential oils: A review. *Food Res. Int.*, 45: 722–734.
- [Banu J](#) and [Gayathri V](#) (2016): Preparation of Antibacterial Herbal Mouthwash against Oral Pathogens. *Int. J.Curr.Microbiol.App.Sci.*, 5(11):205-221
- [Barros JC](#), [Conceição ML](#), [Gomes Neto NJ](#), [Costa ACV](#), [Siqueira Júnior JP](#) and [Basílio Júnior ID](#) (2009): Interference of *Origanum vulgare* L. essential oil on the growth and some physiological characteristics of *Staphylococcus aureus* strains isolated from foods. *LWT Food Sci. Technol.*, 42:1139–1143.
- [Bassolé IHN](#), [Lamien-Meda A](#), [Bayala B](#), [Tirogo S](#), [Franz C](#), [Novak J](#), [Nebié RC](#) and [Dicko MH](#) (2010): Composition and antimicrobial activities of *Lippia multiflora* Moldenke, *Mentha x piperita* L. and *Ocimum basilicum* L. essential oils and their major monoterpene alcohols alone and in combination. *Molecules*, 15:7825–7839.
- [Becheker I](#), [Becheker A](#), [Melakhessou MA](#), [Marref SE](#) and [Berredjem H](#) (2022): Antibacterial, Antifungal, Cytotoxic and Genotoxic Activities of Different Extracts of Arabic and Myrrh Gums. *International Journal of Pharmaceutical Investigation*, 12 (1).
- [Beheshti-Rouy M](#), [Azarsina M](#), [Rezaie-Soufi L](#), [Alikhani MY](#), [Roshanaie G](#) and [Komak S](#) (2015): The antibacterial effect of sage extract (*Salvia officinalis*) mouthwash against *Streptococcus mutans* in dental plaque: a randomized clinical trial. *Iran J Microbiol.* 7(3): 173–177.
- [Ben Arfa A](#), [Combes S](#), [Preziosi-Belloy, L](#), [Gontard N](#) and [Chalier P](#) (2006): Antimicrobial activity of carvacrol related to its chemical structure. *J. Appl. Microbiol.*, 43, 149–154.

- **BHMA, British Herbal Medicine Association (1996):** The British Herbal Pharmacopoeia. Surrey, UK: BHMA.
- **Blumenthal M, Goldberg A and Brinckman J (2000):** Herbal Medicine: Expanded Commission E Monographs. Austin, TX: American Botanical Council.
- **Blumenthal M, Hall T, Goldberg A, Kunz T, Dinda K, Brinkmann J and Keneth J. (2003):** The ABC Clinical Guide to Herbs. American Botanical Council, Austin 51-60.
- **Bornstein MM, Kislig K, Hoti BB, Seemann R and Lussi A (2009):** Prevalence of halitosis in the population of the city of Bern, Switzerland: a study comparing self-reported and clinical data. *Eur J Oral Sci.*, 117(3):261-7.
- **Bornstein MM, Stocker BL, Seemann R, Bürgin WB and Lussi A (2009):** Prevalence of halitosis in young male adults: a study in swiss army recruits comparing self-reported and clinical data. *J Periodontol.*, 80(1):24-31.
- **Boruga O, Jianu C, Mișcă C, Goleț I, Gruia AT and Horhat FG (2014):** Thymus vulgaris essential oil: chemical composition and antimicrobial activity. *Journal of Medicine and Life*; 7(3): 56-60
- **Brieskorn C.H, Noble P, Margaris N, Koedam A and Vokou D (1982):** The Terpenes of the Essential Oil of Myrrh. In: (eds) *Aromatic Plants*. World Crops: Production, Utilization, and Description,7. Springer, Dordrecht. https://doi.org/10.1007/978-94-009-7642-9_19
- **CHEN Y, ZHOU C, GE Z , LIU Y , LIU Y , WEIYI FENG , LI S , CHEN G and WEI T (2013):** Composition and potential anticancer activities of essential oils obtained from myrrh and frankincense. *ONCOLOGY LETTERS*, 6: 1140-1146.
- **Cheng YW, Cheah KP, Lin CW, Li JS, Yu WY, Chang ML, Yeh GC, Chen SH, Choy CS and Hu CM (2011):** Myrrh mediates haem oxygenase-1 expression to suppress the lipopolysaccharide-induced inflammatory response in RAW264.7 macrophages. *J Pharm Pharmacol*, 63:1211–1218.
- **Committee on Herbal Medicinal Products (2013):** Assessment report on *Thymus vulgaris L.*, vulgaris zygis L., herba. European Medicines Agency.

- **Craft J D, Satyal P and Setzer WN (2017):** The Chemotaxonomy of Common Sage (*Salvia officinalis*) Based on the Volatile Constituents. *Medicines (Basel)*, (3):47.
- **D'Amelio FS and Mirhom YW(2010):** Method and composition for treating oral bacteria and inflammation. *io-Botanica, Inc.*, Hauppauge, NY (US) United States Patent Nov.9.
- **Danial EN and Majrashi DM (2016):** Potential scavenging, antimicrobial activities and total phenolic content achieved by resins for five species of *Commiphora*. *Int. J. Adv. Res. Biol. Sci.*, 3(11): 53-61.
- **De Rapper S, Van Vuuren SF, Kamatou GP, Viljoen AM and Dagne E (2012):** The additive and synergistic antimicrobial effects of select frankincense and myrrh oils—a combination from the pharaonic pharmacopoeia. *Letters in applied microbiology*, 54(4):352–8.
- **Dolara P, Luceri C, Ghelardini C et al. (1996):** Analgesic effects of myrrh. *Nature*, 376: 29.
- **Dorman HJD and Deans SG (2000):** Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *Journal of Applied Microbiology*, 88(2): 308-316.
- **ESCOP (1996):** *Salviae folium* (Sage leaf). Monographs on the Medicinal Use of Plant Drugs. Exeter, UK: European Scientific. Cooperative on Phytotherapy.
- **Faveri M, Hayacibara MF, Pupio GC, Cury JA, Tsuzuki CO and Hayacibara RM (2006):** A cross-over study on the effect of various therapeutic approaches to morning breath odour. *J Clin Periodontol*, 33(8):555-60.
- **Fedorowicz Z, Aljufairi H, Nasser M, Outhouse TL and Pedrazzi V (2008):** Mouthrinses for the treatment of halitosis. *Cochrane Database Syst Rev.*, 8(4):CD006701.
- **Ferrazzano GF, Roberto L, Amato I, Cantile T, Sangianantoni G, and Ingenito A (2011):** Antimicrobial Properties of Green Tea Extract Against Cariogenic Microflora: An In Vivo Study. *J Med Food*, 14(9):907-11.
- **Gali-Muhtasib H, Hilan C and Khater C (2000):** Traditional uses of *Salvia libanotica* (East Mediterranean sage) and the effects of its essential oils. *Journal of Ethnopharmacology*, 71 : 513-520.

- **Gill AO, Delaquis P, Russo P and Holley RA (2002):** Evaluation of antilisterial action of cilantro oil on vacuum packed ham. *International Journal of Food Microbiology*, 73(1): 83-92.
- **Goncalves GMS, Bottaro M and Nilson F (2011):** Effect of the Thymus vulgaris essential oil on the growth of *Streptococcus mutans*. *Journal of basic and applied pharmaceutical sciences*, 32(3):375-380.
- **Graham HN (1992):** Green tea composition, consumption, and polyphenol chemistry. *Prev Med.*; 21(3):334-50.
- **Gultz J, Kaim JM., DeLeo J and Sherer W (1998):** An *in vivo* comparison of the antimicrobial activities of three mouthrinses. *J. Clin. Dent.*, 9: 43-45.
- **Hazzit M, Baaliouamer A, Verissimo AR, Falerio ML and Miguel MG (2009):** Chemical composition and biological activities of Algerian thymus oils. *Food Chem.*, 116, 714–721.
- **Hose S. (2002):** Der Wermut-Artemisia absinthium L. *Zeitschrift Phytother*, 23: 187-194.
- **Hu C, Liang Y, Guo F, Li X and Wang W (2010):** Determination of Essential Oil Composition from *Osmanthus fragrans* Tea by GC-MS Combined with a Chemometric Resolution Method. *Molecules*, 15: 3683-3693.
- **Jirovetz L, Buchbauer G, Denkova Z, Slavchev A, Stoyanova A and Schmidt E (2006):** Chemical composition, antimicrobial activities and odor descriptions of various *Salvia* sp. and *Thuja* sp. essential oils. *Ernährung/Nutrition*, 30: 152–159.
- **Kahkönen MP, Hopia AI, Vuorela HJ, Rauha JP, Pihlaja K, Ku-jala TS and Heinonen M (1999):** Antioxidant activity of plant extracts containing phenolic compounds. *J Agric Food Chem*;47:3954–3962.
- **Kaneko K, Shimano N, Suzuki Y, Nakamukai M, Ikazaki R, Ishida N, Kanayasu E, Karuda T, Takihara T, Sakane I, Yayabe F and Matsui T (1993):** Effects of tea catechins on oral odor and dental plaque. *Oral Therapeutics and Pharmacology*.;12:189–197.
- **Kawamura J and Takeo T (1989):** Antibacterial activity of tea catechin to *Streptococcus mutans*. *Journal of the Japanese Society of Food Science and Technology*.;36:463–467.

- **Kazor CE, Mitchell PM, Lee AM, Stokes LN, Loesche WJ, Dewhirst FE, et al. (2003):** Diversity of bacterial populations on the tongue dorsa of patients with halitosis and healthy patients. *J Clin Microbiol.*, 41(2):558-63.
- **Komes D, Horžić D, Belščak A, Ganić KK and Vulić I (2010):** Green Tea Preparation and Its Influence on the Content of Bioactive Compounds. *Food Res. Int.*, 43, 167–176.
- **Kubo I, Muroi H and Himejima M (1992):** Antimicrobial activity of green tea flavor components and their combination effects. *Journal of Agricultural and Food Chemistry.*;40:245–248.
- **[Kumar G](#), [Jalaluddin MD](#), [Rout P](#), [Mohanty R](#) and [Dileep CL \(2013\):](#) Emerging Trends of Herbal Care in Dentistry. *J Clin Diagn Res.*, 7(8): 1827–1829.**
- **Kumari R, Meyyappan A, Nandi D, Agrawalla BK, Chowdhury AA, Selvamani P, Latha S, Giri VS, Mukherjee J, Bandopadhyay S and Jaisankar P (2011) :** Antioxidant and antibacterial activities of bark extracts from *Commiphora berryi* and *Commiphora caudate*. *Nat Prod Res.*, 25:1454–1462.
- **MAHBOUBI M and KAZEMPOUR N (2016):** The antimicrobial and antioxidant activities of *Commiphora molmol* extracts. *BIHAREAN BIOLOGIST*, 10 (2): 131-133
- **Marino M and Bersani CC (1999):** Antimicrobial Activity of the Essential Oils of *Thymus vulgaris* L. Measured Using a Bioimpedometri Method. *J Food Prot.*, 62: 1017-23.
- **Massoud A, El Sisi S, Salama O and Massoud A (2001):** Preliminary study of therapeutic efficacy of a new fasciolicidal drug derived from *Commiphora molmol* (Myrrh). *Am J Trop Med Hyg*, 65:96–99.
- **Matalon S, Weiss EI, Gorfil C, Noy D and Slutzky H (2009):** In vitro antibacterial evaluation of flowable restorative materials. *Quintessence Int.*, 40: 327– 332.
- **McCormick, K., and Salcedo, J. (2017).** SPSS statistics for data analysis and visualization. John Wiley & Sons.
- **MITIĆ-ĆULAFIĆ D, VUKOVIĆ-GAČIĆ B, KNEŽEVIĆ-VUKČEVIĆ J, STANKOVIĆ S and SIMIĆ D (2005):** Comparative study on the antibacterial

activity of volatiles from sage (*Salvia officinalis*L.). *Arch. Biol. Sci.*, Belgrade, 57 (3): 173-178.

- **Miyazaki H, Sakao S, Katoh Y and Takehara T (1995):** Correlation between volatile sulphur compounds and certain oral health measurements in the general population. *J Periodontol*, 66(8):679-84.
- **Mohamed A, Mohamed A and Omar AA(2013):** A study to find thyme oil dose that kill 50% of mice and minimal dose that kill all mice and maximum nonlethal Dose. *Nature and Science.*, 11(12): 52-53.
- **[Neturi RS](#), [Srinivas R](#), [Simha VB](#), [Sree SY](#), [Shekar CT](#) and [Kumar P S](#) (2014):** Effects of Green Tea on *Streptococcus mutans* Counts- A Randomised Control Trail. *J Clin Diagn Res.*, 8(11): ZC128–ZC130
- **Nissen HP, Blitz H, Kreyel HW(1988):** Prolifometrie, eine methode zur beurteilung der therapeutischen wirksamkeit kon Kamillosan_-Salbe. *Z Hautkr*, 63: 184–190.
- **Orav, A; Raal, A and Arak, E (2010):** Content and composition of the essential oil of Chamomilla recutita (L.) Rauschert from some European countries. *Natural Product Research*, 24: 48–55.
- **Palombo EA (2011):** Traditional medicinal plant extracts and natural products with activity against oral bacteria: potential application in the prevention and treatment of oral diseases. *Evid Based Complement Altern Med* 2011:E680354
- **Pastoriza S, Mesías M, Cabrera C and Rufián-Henares JA (2017):** Healthy Properties of Green and White Teas: An Update. *Food Funct.*, 8, 2650–2662.
- **Pistorius A, Willershausen B, Steinmeier EM and Kreisler M. (2003):** Efficacy of subgingival irrigation using herbal extracts on gingival inflammation. *J. Periodontal*, 74: 616-22.
- **Pourabbas R, Delazar A and Chitsaz MT (2005):** The effect of German chamomile mouthwash on dental plaque and gingival inflammation. *Iranian Journal of Pharmaceutical Research.*,2:105–9.
- **Quirynen M, Dadamio J, Van den Velde S, De Smit M, Dekeyser C, Van Tornout M, et al. (2009):** Characteristics of 2000 patients who visited a halitosis clinic. *J Clin Periodontol.*, 36(11):970-5.
- **Raal, A; Orav, A; Püssa, T; Valner, C; Malmiste, B and Arak, E (2012):** Content of essential oil, terpenoids and polyphenols in commercial chamomile

(*Chamomilla recutita* L. Rauschert) teas from different countries. *Food Chemistry*, 131 (2):632-638.

- **Rahman M, Garvey M, Laura JVP and Simon G (2008):** Antibacterial terpenes from the oleo-resin of *Commiphora molmol* (Engl.). *Phytother Res.*, 22:1356–1360.
- **Rasheed A and Haider M (1998):** Antibacterial activity of *Camellia sinensis* extracts against dental caries. *Arch Pharm Res.*, 21(3):348-52.
- **Rösing CK and Loesche W (2011):** Halitosis: an overview of epidemiology, etiology and clinical management. *Braz. oral res.*, 25 (5) São Paulo.
- **Rota MC, Herrera A, Martínez RM, Sotomayor JA and Jordán MJ (2008):** Antimicrobial activity and chemical composition of *Thymus vulgaris*, *Thymus zygis* and *Thymus hyemalis* essential oils. *Food Control*. 19(7): 681-687.
- **Sağdıç O, Kuşçu A, Özcan M and Özçelik S (2002):** Effects of Turkish spice extracts at various concentrations on the growth of *Escherichia coli* O157:H7. *Food Microbiology*, 19(5): 473-480.
- **Sakanaka S, Kim M, Taniguchi M and Yamamoto T (1989):** Antibacterial substances in Japanese green tea extract against *Streptococcus mutans*, a cariogenic bacterium. *Agricultural and Biological Chemistry.*;53:2307–2311.
- **Salako NO and Philip L (2011):** Comparison of the use of the Halimeter and the Oral Chroma[®] in the assessment of the ability of common cultivable oral anaerobic bacteria to produce malodorous volatile sulfur compounds from cysteine and methionine. *Med Princ Pract.*, 20(1):75-9.
- **Seidler-Łożykowska K, Mordalski R, Król D, Bocianowski J and Karpińska, E (2015):** Yield and quality of sage herb (*Salvia officinalis* L.) from organic cultivation. *Biol. Agric. Hortic.*, 31: 53–60.
- **Selim S, Almuhayawi MS, Alqhtani H, Al Jaouni, SK, Saleh MW, Warrad M and Hagagy N (2022):** Anti-Salmonella and Antibiofilm Potency of *Salvia officinalis* L. Essential Oil against Antibiotic-Resistant *Salmonella enterica*. *Antibiotics*, 11(4): 489.

- **Shapiro S and Guggenheim (1995)**: The action of thymol on oral bacteria. *Oral microbiology*, 10 (4):241-246.
- **Shapiro S, Meier A and Guggenheim B (1994)**: The antimicrobial activity of essential oils and essential oil components towards oral bacteria. *Oral microbiology*, 9 (4): 202-208.
- **Sheir Z, Nasr AA, Massoud A et al. (2001)**: A safe, effective, herbal antischistosomal therapy derived from myrrh. *Am J Trop Med Hyg*, 65: 700–704.
- **Shin JY, Che DN, Cho BO, Kang HJ, Kim J and Jang SI (2019)**: Commiphora myrrha inhibits itch-associated histamine and IL-31 production in stimulated mast cells. *Experimental and therapeutic medicine*, 1;18(3):1914–20.
- **Shinada K, Ueno M, Konishi C, Takehara S, Yokoyama S, Zaitzu T, et al. (2010)**: Effects of a mouthwash with chlorine dioxide on oral malodor and salivary bacteria: a randomized placebo-controlled 7-day trial. *Trials*. 2010 Feb 12;11:14
- **Silveira EMV, Piccinin FB, Gomes SC, Oppermann RV and Rösing CK (2011)**: The effect of gingivitis treatment on the breath of chronic periodontitis patients. *Oral Health Prev Dent. Forthcoming*.
- **Singh O, Khanam Z, Misra N and Srivastava MK (2011)**: Chamomile (*Matricaria chamomilla* L.): An overview. *Pharmacogn. Rev.*, 5, 82–95.
- **Singh, G; Maurya, S; deLampasona, M P and Catalan, C A N (2007)**: A comparison of chemical, antioxidant and antimicrobial studies of cinnamon leaf and bark volatile oils, oleoresins and their constituents. *Food and Chemical Toxicology*, 45: 1650–1661.
- **Söder B, Johansson B and Söder PO (2002)**: The relation between foetor ore, oral hygiene and periodontal disease. *Swed Dent J.* , 24(3):73-82.
- **Soković MD, Vukojević J, Marin PD, Brkić DD Vajs, V and van Griensven L.J.L.D (2009)**: Chemical composition of essential oils of *Thymus* and *Mentha* species and their antifungal activities. *Molecules*, 14, 238–249.
- **Stahl-Biskup (1989)**: The Chemical Composition of *Thymus* Oils: A Review of the Literature 1960–1989. *Journal of Essential Oil Research*, 3(2).
- **Takeshita T, Suzuki N, Nakano Y, Shimazaki Y, Yoneda M, Hirofuji T, et al (2010)**: Relationship between oral malodor and the global composition of

indigenous bacterial populations in saliva. *Appl Environ Microbiol.*, 76(9):2806-14.

- **Taylor, PW, Hamilton-Miller, JMT and Stapleton PD (2005):** Antimicrobial properties of green tea catechins. *Food Sci Technol Bull.*, 2: 71–81.
- **Treadway S (1998):** Exploring the universe of Ayurvedic botanicals to manage bacterial infections. *Clin Nutr Insight*, 6:1–3.
- **Van Wyk BE and Wink M (2004):** Medicinal plants of the world an illustrated scientific guide to important medicinal plants and their uses. Timber Press, Portland.
- **Viuda-Martos M, Navajas YR, Zapata ES, Fernández-López J and Pérez-Álvarez JA (2010):** Antioxidant activity of essential oils of five spice plants widely used in a Mediterranean diet. *Flavour Fragr. J.*, 25: 13–19.
- **Waskmundzka M, Wianowska D, Szewczyk, K and Oniszczuk A (2007):** Effect of samplepreparation methods on the HPLC quantitation of some phenolic acids in plant materials. *Acta Chromatographica*, (19): 227-237.
- **Weiss RF (1988):** Herbal Medicine. Gothenburg, Sweden: Ab Arcanum and Beaconsfield, UK: *Beaconsfield Publishers L*, 208–209.
- **Yasuda H and Arakawa T (1995):** Deodorizing mechanism of (–)-epigallocatechin against methyl mercaptan. *Bioscience, Biotechnology and Biochemistry.*;59:1232–1236.
- **Yokoyama S, Ohnuki M, Shinada K, Ueno M, Wright FA and Kawaguchi Y (2010):** Oral malodor and related factors in Japanese senior high school students. *J Sch Health.*, 80(7):346-52.
- **Yoneda M, Masuo Y, Suzuki N, Iwamoto T and Hirofuji T (2010):** Relationship between the β -galactosidase activity in saliva and parameters associated with oral malodor. *J Breath Res.*, 4(1):17-8.

تأثير بعض الزيوت العطرية علي بعض بكتريا الفم الممرضة للإنسان

مني ياسر عبد الخالق مصطفى**

أشرف رفعت محمد الزيني*

الملخص العربي:

من المعروف أن صحة وسلامة المعدة مرتبطة بصحة وسلامة الفم، لذا تهدف هذه الدراسة الي اختبار فاعلية زيوت الأعشاب المختارة ، على بكتيريا الفم لمحتواها من الزيوت العطرية. حيث تم دراسة النشاط المضاد للبكتيريا من نبات المر (*Commiphora molmol*) والزعتر (*Thymus vulgaris L*) والمريمية (*Salvia officinalis L*) والبابونج (*Matricaria recutita L*) والشاي الأخضر (*Camellia sinensis*)، تمت دراسة تأثير الزيوت الأساسية بتركيز (٥ مجم زيت / ١٠٠ مل ماء) على العدد البكتيري الكلي في اللعاب، كما تم دراسة تأثير تلك الزيوت بذات التركيز علي العدد الكلي لكلاً من *Lactobacillus rhamnosus* و *Streptococcus mutans* وكذلك الحد الأدنى من المناطق المثبطة (MIZ) من زيوت نبات المر والزعتر والمريمية والبابونج والشاي الأخضر على *Lactobacillus rhamnosus* و *Streptococcus mutans* باستخدام تقنية أجار الحساسية القياسية. حيث تمت معالجة البيانات إحصائياً، وأظهرت المجموعة التجريبية انخفاضاً ذا دلالة إحصائية في تعداد مستعمرات *Lactobacillus rhamnosus* و *Streptococcus mutans* بالنسبة للمجموعة المقارنة. أظهرت هذه النتائج فعالية زيوت الأعشاب المختبرة ضد رائحة الفم الكريهة والفلورا الفموية ، وتحقيق توازن بين بكتيريا الفم الضارة وغير الضارة ، وكذلك فتح طريق واعد للتطبيقات السريرية في تحضير علاجات محددة وطبيعية مضادة للبكتيريا. توصي الدراسة باستخدام زيوت الأعشاب موقع الدراسة بالتركيز المذكور في السيطرة علي بكتريا الفم الممرضة حفاظا علي صحة المعدة والجسم.

* أستاذ مساعد - قسم علوم الأغذية - كلية الزراعة - جامعة القاهرة - الجيزة- مصر
** زميل تعليمي تخصص تغذية وعلوم الأطعمة بالهيئة العامة للمستشفيات والمعاهد التعليمية - مستشفى الجلاء التعليمي